Tetrahedron Letters No.22, pp. 2459-2463, 1966. Pergamon Press Ltd. Printed in Great Britain.

A NEW C₂₀ GIBBERELLIN IN IMMATURE SEEDS OF <u>LUPINUS</u> <u>LUTEUS</u> Koichi Koshimizu, Hiroshi Fukui, Toshiatsu Kusaki, Tetsuo Mitsui and Yukiyoshi Ogawa* Department of Agricultural Chemistry, Kyoto University, Kyoto, Japan. (Received 22 March 1966)

Extracts of immature seeds of <u>Lupinus luteus</u> have been found to possess biological activities similar to the gibberellins (1-4). It was suggested from the chromatographic behavior and the characteristics of infrared spectra that the active principles were different from C_{19} gibberellins (5). We wish to report the isolation of a new gibberellin, "Lupinus gibberellin-I", from the seeds, and to propose the structure shown as I for the gibberellin.

The method of isolation followed that of MacMillan et al. (6). The fraction eluted from the charcoal-celite column with water containing 59-62 % acetone was rechromatographed on silica-celite. Elution with 50-55 % ethyl acetate in chloroform yielded 35 mg. of Lupinus gibberellin, (L_1) , from 60 kg. of the seeds. L_1 , (I), which formed needles from acetone, m.p. 240-242⁰, gave elementary analysis

* Laboratory of Applied Botany, Faculty of Agriculture, Kyoto University. consistent with the formula $C_{20}H_{28}O_6$.

The infrared (I.R.) absorption (nujol) at 3400-3100 br., 2500 br., 1703 and 1673 cm^{-1} , and no ultraviolet absorption between 250 and 350 mu indicated the presence of hydroxyl and carboxyl group in L₁. The I.R. spectrum of the corresponding methyl ester (II) (KBr) showed absorption attributable to hydroxyl (3550-3300 br. cm^{-1}), ester carbonyl (1725 br. cm^{-1}) and terminal methylene (1667 and 894 cm^{-1}). Under normal acetylation conditions, the methyl ester formed a monoacetate (III) which had I.R. absorption (CHCl_{τ}) at 3585 (tertiary hydroxyl), 1720 br. (ester carbonyl), 1665 and 896 ${\rm cm}^{-1}$ (terminal methylene), and had nuclear magnetic resonance (N.M.R.) signals at 0.70 and 0.97 δ (two tertiary methyls), 2.07 δ (acetate), 3.67 and 3.69 δ (two carbomethoxyls), 4.87 and 5.10 δ (terminal methylene, multiplet) and 5.29 δ (H-C-OAc, triplet, J=2.5 c.p.s.). In addition, a pair of doublets centered at 2.26 and 3.26 & (J=12 c.p.s.) was assigned to the 10, 10a AB-quartet which is a characteristic feature of the gibbane skeleton in the known gibberellins and their derivatives (7-9). The presence of a tertiary hydroxyl proton signal at 2.11 δ was confirmed by shaking the solution of the sample with deuterium oxide. L,,(I), has therefore five pendant carbon groups on a gibbane ring as GA_{12} (10) and GA_{13} (11) which were isolated from fermentations of Gibberella fujikuroi.

Treatment of L_1 with dilute hydrochloric acid at 100° gave a keto-acid (IV), and methylation of the keto-acid



produced a dimethyl ester which readily formed a monoacetate (V). The resultant monoacetate (V) showed the I.R. absorption band (CCl₄) at 1740 br. cm⁻¹, and the N.M.R. signals at 0.67, 0.97 and 0.98 & (three tertiary methyls), 2.10 & (acetate), 2.40 (doublet, J=12 c.p.s.) and 3.38 & (doublet, J=12 c.p.s.) (10, 10a protons), 3.69 & (two carbomethoxyls) and 5.32 & (H-¢-OAc, triplet, J=2.5 c.p.s.). Comparison of the spectra of III and V showed that the acid treatment caused the rearrangement which is analogous to the Wagner-Meerwein rearrangement of GA₁ (12, 13), GA₅ (14), GA₆ and GA₈ (6) to their keto-acids.

The presence of a gibbane AB-quartet in L_1 derivatives required a carboxyl group at C-10, and the rearrangement of L_1 ; (I), to the keto-acid (IV) suggested that the tertiary hydroxyl group was placed at C-7 and the terminal methylene group at C-8. The keto-acid (IV) on treatment with the Jones reagent was oxidized to a diketo-acid which was decarboxylated at 100° to give a diketo-monoacid (VI). The N.M.R. spectrum of the corresponding methyl ester (VII) showed signals at 0.83 δ (=CH-CH₃, doublet, J=6 c.p.s.), 0.98 and 1.02 δ (two tertiary methyls) and 3.67 δ (carbomethoxyl). The hydroxyl group was thus placed at C-2 which was adjacent to C-1 bearing both a tertiary methyl group and a carboxyl group.

Lupinus gibberellin-I was assayed on rice seedlings (15). It increased the second leaf sheath length to ca. 200 % of the control at an optimum concentration of 10 mg./1.

The N.M.R. spectra were measured on a Varian A6O spectrometer in carbontetrachloride with tetramethylsilane as internal reference.

We thank Dr. T. Shingu for the measurements of N.M.R. spectra and Prof. M. Nakajima for the I.R. absorption spectra.

REFERENCES

1.	Y٠	Murakami,	Bot.	Mag.	(Tokyo)	, 72,	438	(1959)).

- Y. Ogawa and S. Imamura, <u>Bot. Mag. (Tokyo</u>), <u>73</u>, 125 (1960).
- Y. Hirono, Y. Ogawa and S. Imamura, <u>Plant & Cell Physiol.</u>, <u>1</u>, 81 (1960).
- 4. Y. Ogawa, Plant & Cell Physiol., 4, 85 (1963).
- 5. Y. Ogawa, Bot. Mag. (Tokyo), in impress.

- 6. J. MacMillan, J.C. Seaton and P.J. Suter, <u>Tetrahedron</u>, <u>18</u>, 349 (1962).
- 7. N. Sheppard, <u>J. Chem. Soc</u>., 3040 (1960).
- D.C. Akdridge, J.F. Grove, R.N. Speake, B.K. Tidd and W. Klyne, <u>J. Chem. Soc</u>., 143 (1963).
- 9. J.R. Hanson, J. Chem. Soc., 5036 (1965).
- 10. B.E. Cross and K. Norton, <u>J. Chem. Soc</u>., 1570 (1965).
- 11. R.H.B. Galt, J. Chem. Soc., 3143 (1965).
- N. Takahashi, Y. Seta, H. Kitamura, A. Kawarada and Y. Sumiki, <u>Bull. Agric. Chem. Soc. Japan</u>, <u>21</u>, 75 (1957).
- 13. B.E. Cross, J. Chem. Soc., 3022 (1960).
- 14. J. MacMillan, J.C. Seaton and P.J. Suter, <u>Tetrahedron</u>, <u>11</u>, 60 (1960).
- 15. Y. Ogawa, Plant & Cell Physiol., 4, 227 (1963).